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Novel potent and selective calcium-release-activated calcium (CRAC) channel inhibitors. Part 1: Synthesis and inhibitory activity of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides

Yasuhiro Yonetoku,* Hirokazu Kubota, Yoshinori Okamoto, Akira Toyoshima,[†]
Masashi Funatsu, Jun Ishikawa, Makoto Takeuchi,
Mitsuaki Ohta and Shin-ichi Tsukamoto

Drug Discovery Research, Astellas Pharma Inc., 21 Miyukigaoka, Tsukuba, Ibaraki 305-8585, Japan
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Abstract—We synthesized and evaluated a series of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides to identify potent inhibitors of calcium-release-activated calcium (CRAC) channels with greater selectivity than voltage-operated calcium (VOC) channels. These efforts resulted in identification of compounds 22 and 24. The former exhibits highly potent and selective CRAC channel inhibitory activity, and the latter inhibited phytohemagglutinin-induced interleukin-2 production by Jurkat T lymphocytes and concanavalin A-induced hepatitis in mice.

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1. Introduction

Ca²⁺ channels in the plasma membrane are known to regulate Ca2+ influx, and as a result they regulate a variety of cellular functions. Voltage-operated calcium (VOC) channels, located in the nervous, endocrine, cardiovascular, and skeletal systems, are known to be modulated by the membrane potential, and are classified into L, N, P, Q, R, and T subtypes. Some of these molecules have been well characterized, and their inhibitors, such as nifedipine, have been used for the treatment of hypertension¹⁶ and brain dysfunction.^{1c} In contrast, Ca²⁺ channels in inflammatory cells, such as lymphocytes, mast cells and neutrophils, are activated regardless of their membrane potential^{2a} and are known as storeoperated Ca²⁺ channels. They have been shown to play important roles in the pathogenesis and exacerbation of inflammatory and autoimmune diseases.2b-e

The early stages in T lymphocyte activation can be divided into events pre- and post-intracellular Ca²⁺ release.³ Before Ca²⁺ release, the stimulation of cell surface receptors induces the generation of IP₃, followed by the release of Ca²⁺ from the endoplasmic reticulum (ER).⁴ After the depletion of Ca²⁺ from ER stores, calcium-release-activated calcium (CRAC) channels are activated and capacitative Ca²⁺ influx through these channels sustains a high intracellular Ca²⁺ concentration ([Ca²⁺]_i).⁵ This prolonged high [Ca²⁺]_i is crucial for cytosolic signal transduction which induces the production of lipid mediators (e.g., LTD₄), cytokines (e.g., interleukin-2 (IL-2)), and matrix metalloproteinases, which are known to play important roles in the pathogenesis of inflammation and autoimmune diseases.

These facts suggested that CRAC channel inhibitors would be useful for the treatment of diseases, such as asthma and rheumatoid arthritis, caused by the activation of inflammatory cells. However, inhibitors which affect both CRAC and VOC channels are predicted to show undesirable side-effects in the nervous and cardiovascular systems. Therefore, it may be necessary for CRAC channel inhibitors to exhibit sufficient selectivity over VOC channels to be useful as anti-inflammatory drugs.

Keywords: Anti-inflammatory; Channel inhibitors; Ca²⁺-release-activated Ca²⁺ channel; CRAC channel; Interleukin-2; Quantitative structure-activity relationship.

^{*} Corresponding author. Tel.: +81 0 29 847 8611; fax: +81 0 29 847 8313; e-mail: yasuhiro.yonetoku@jp.astellas.com

[†] Present address: Development Division, Astellas Pharma Inc., 3-17-1 Hasune, Itabashi, Tokyo 174-8612, Japan

In the last decade, several compounds including SK&F 96365 (1),⁷ LOE 908 (2),⁸ and L-651582 (3,⁹ Fig. 1) have been reported to inhibit CRAC channels. However, their potency and selectivity are unlikely to be adequate for their use as anti-inflammatory drugs. To find potent and selective CRAC channel inhibitors, we screened our chemical library and identified *N*-(4-chlorophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (4,¹⁰ Fig. 1) as a lead compound. We have studied the structure–activity relationships (SARs) of this compound and in this paper, we describe the synthesis of a series of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides, and the SARs of their 4-chlorophenyl moiety.

2. Chemistry

The 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides were prepared as shown in

Schemes 1–3. In the presence of sodium methoxide (NaOMe) and ethyl trifluoroacetate, 2-acetylthiophene 5 was converted to the dione 6,11 and regioselective cyclization of compound 6 with methylhydrazine produced the pyrazole 7 (Scheme 1). The structure of compound 7 was determined as illustrated in Schemel, using the nuclear Overhauser effect (NOE), observed between the protons of the N-methyl group (4.01 ppm) and the thiophene (7.21 ppm). Lithiation of compound 7 with n-butyllithium (n-BuLi) in tetrahydrofuran (THF), followed by treatment with ethyl chloroformate, gave the ester 8, which was hydrolyzed with aqueous NaOH to form the acid 9. Compound 9 was treated with oxalyl chloride and the resultant acyl chloride 10 was allowed react with amines to form 5-(1-methyl-3trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides (11–35). The 4-hydroxyphenyl (36) and the 1-*tert*-butoxycarbonylpiperidin-4-yl (37) derivatives were prepared by the reaction of compound 9 with the corresponding amines¹² in the presence of 1-ethyl-3-(3-

Figure 1. Structures of SK&F 96365 (1), LOE 908 (2), L-651582 (3), and compound 4.

Scheme 1. Reagents: (a) CF₃CO₂Et, NaOMe, MeOH; (b) MeNHNH₂, AcOH–EtOH; (c) *n*-BuLi, THF, then ClCO₂Et; (d) 1 M NaOH, EtOH; (e) (COCl)₂, cat. DMF, THF; (f) R-NH₂, base; (g) R-NH₂, EDC·HCl, THF.

Scheme 2. Reagents: (a) Zn, NH₄Cl, EtOH–H₂O.

Scheme 3. Reagents: (a) 4 M HCl (g), AcOEt.

Table 1. Biological properties of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(4-substituted-phenyl)-2-thiophenecarboxamides

Compound	R	IC ₅₀ (μM)		CRAC channel selectivity index ^c
		CRAC ^a	VOCb	
4	Cl	0.13	0.75	5.8
11	F	0.14	3.6	26
12	Br	0.27	2.6	10
13	CO_2Et	46% ^d	NTe	_
14	CN	0.56	1.5	2.7
15	NO_2	1.4	4.0	2.9
16	Н	0.062	1.9	31
17	Me	0.37	2.4	6.5
18	<i>i</i> Pr	0.92	1.2	1.3
36	OH	0.24	4.3	18
19	OMe	0.44	$24\%^{f}$	>23 ^g
38	NH_2	4.1	$69\%^{f}$	_
20	NMe_2	8% ^f	NT	_

^a Inhibition of Ca²⁺ influx through CRAC channels on Jurkat T lymphocytes. See experimental section.

dimethylaminopropyl)carbodiimide hydrochloride (EDC·HCl), as shown in Scheme 1. Reduction of the 4-nitrophenyl derivative 15 with zinc powder gave the 4-aminophenyl derivative 38 (Scheme 2), and the piperidin-4-yl derivative 39 was obtained from compound 37 by treatment with HCl (Scheme 1).

3. Results and discussion

The compounds synthesized as described above were evaluated for their ability to inhibit Ca²⁺ influx through CRAC channels on Jurkat T lymphocytes¹³ and VOC

channels on PC12-h5 cells. ¹⁴ The results are summarized in Tables 1–3.

We first examined the effects of substitutions at the 4-position in the phenyl group of compound 4 on the inhibitory activity and selectivity (Table 1). Substituting fluoro (11) and bromo (12) groups for the chloro group did not change the compounds' inhibitory effects on CRAC channels, but reduced their inhibitory activity against VOC channels. Substituting ethoxycarbonyl (13), cyano (14), and nitro (15) groups reduced the inhibitory effect on CRAC channels, suggesting that substituting groups that were highly electron-withdrawing at the 4-position would not favor increased CRAC channel inhibitory activity. In contrast, removal of the chloro group significantly increased CRAC channel inhibitory activity, with compound 16 showing an IC₅₀ value of 0.062 μM, and in addition exhibiting high selectivity for CRAC channels with an index of over 31. The methyl derivative 17 was almost equipotent to compound 4, whereas the isopropyl derivative 18 was a less effective inhibitor. From these results, we speculated that the ability to inhibit CRAC channels was sensitive to the bulkiness of the group at the 4-position. Hydroxy (36) and methoxy (19) groups, which maintained the potency, improved the selectivity for CRAC channels, with indices of approximately 20. In contrast, substituting nitrogen-containing groups such as amino (38) and dimethylamino (20) groups at this position dramatically decreased the inhibitory activity.

To investigate the SARs in detail, we performed a quantitative analysis using various parameters for side groups, such as π , ¹⁵ molecular refractivity, ¹⁶ Swein

Table 2. Biological properties of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(substituted-phenyl)-2-thiophenecarboxamides

Compound	R	IC ₅₀ (μM)		CRAC channel
		CRAC ^a	VOC ^b	selectivity index ^c
4	4-Cl	0.13	0.75	5.8
21	3-C1	1.1	2.2	2.0
22	2-C1	0.050	$0\%^{d}$	>200 ^e
23	2-F	0.090	4.0	44
24	2-Me	0.30	10	33
25	2,4-diCl	0.14	18% ^d	>71 ^e

^{a-c}See the corresponding footnotes to Table 1.

^b Inhibition of Ca²⁺ influx through VOC channels on PC12-h5 cells. See experimental section.

^c IC₅₀ for VOC channels/IC₅₀ for CRAC channels.

 $[^]d$ % inhibition at 0.3 μ M.

e Not tested.

^f% inhibition at 10 μM.

 $^{^{}g}$ IC₅₀ value for VOC channels was taken as >10 μ M.

 $[^]d\,\%$ inhibition at 10 $\mu M.$

 $^{^{}e}\,IC_{50}$ value for VOC channels was taken as >10 $\mu M.$

Table 3. Biological properties of 5-(1-methyl-3-trifluoromethyl-1H-pyrazol-5-yl)-N-substituted-2-thiophenecarboxamides

Compound	R	IC ₅₀ (μM)		CRAC channel selectivity index ^c
		CRAC ^a	VOC^{b}	
4	4-Chlorophenyl	0.13	0.75	5.8
26	Thiophen-2-yl	0.10	1.2	12
27	Thiophen-3-yl	0.61	2.3	3.8
28	1-Methylpyrrol-2-yl	0.58	10	17
29	Pyridin-3-yl	0.99	47% ^d	>10 ^e
30	Pyridin-4-yl	0.42	4.1	10
31	Thiazol-2-yl	$35\%^{\rm f}$	NT^g	_
32	1,2,4-Triazol-4-yl	$6\%^{\mathrm{d}}$	NT^g	_
33	Tetrazol-5-yl	12% ^d	NT^g	_
34	Ethyl	1.7	17% ^d	>6 ^e
35	Cyclohexyl	0.36	4.3	12
39	Piperidin-4-yl	$10\%^{d}$	NT^g	_

^{a-e}See the corresponding footnotes to Table 1.

and Lupton F/R, ¹⁷ $\sigma_{\rm para}$, ¹⁸ Taft $E_{\rm s}$, ¹⁹ Verloop L, ²⁰ molecular volume, molecular surface area, and Verloop B5. ²⁰ For the series of compounds shown in Table 1, the $\sigma_{\rm para}$ values of the group substituted at the 4-position in the phenyl group showed the best correlation (r=0.81) with the ability to inhibit CRAC channels. These results are shown in Figure 2 and described by the following parabolic equation:

$$pIC_{50} = -2.04(\pm 0.44)\sigma_{para}^2 + 6.74$$
 (Eq. 1, $n = 13$, $r = 0.81$, $s = 0.37$, $F = 21.46$).

These results showed that both highly electron-with-drawing and highly electron-donating groups tended to dramatically reduce the inhibitory effect on CRAC channels. On the other hand, compounds containing groups with $\sigma_{\rm para}$ values of around 0 would be expected to act as highly potent inhibitors.

In the series of compounds with chloro groups in the phenyl group (4, 21, and 22), a 3-chloro group (21) decreased both the activity and the selectivity (Table 2). In contrast, the 2-chloro derivative (22), with an

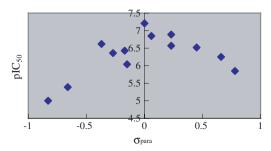


Figure 2. Correlation between Hammett constant (σ_{para}) and pIC_{50} . The pIC_{50} values of compounds **13** and **20** are plotted as 6.5 and 5.0, respectively.

 IC_{50} value of 0.050 μ M, was 3-fold more potent than compound 4 as a CRAC channel inhibitor. Surprisingly, this compound showed no inhibition of VOC channels at 10 μ M, and an excellent selectivity for CRAC channels with an index of more than 200. Other substitutions at the 2-position (23–25) also improved the selectivity of the compounds for CRAC channels, implying that substitutions at this position would generally be expected to reduce inhibitory effects on VOC channels and produce CRAC channel-selective compounds.

Next, we investigated the effects of substituting heteroaromatic and aliphatic groups for the 4-chlorophenyl moiety of compound 4. As shown in Table 3, in the series of compounds containing heteroaromatics, the ability of the compound to inhibit CRAC channels was unaffected by a 2-thienyl group (26), but was reduced 5-fold in the 3-thienyl derivative 27. The 1-methylpyrrolyl (28) and pyridyl (29, 30) derivatives were also less potent than compound 4, and the introduction of thiazolyl (31), 1,2,4-triazolyl (32), and tetrazolyl (33) groups resulted in dramatic reductions in the inhibition of CRAC channels. Among the compounds with aliphatic groups (34, 35, and 39), the cyclohexyl derivative 35 was found to show the most potent CRAC channel inhibitory activity and was almost as potent as compound 4. These results implied that aromatic groups were not essential features of CRAC channel inhibitors.

To investigate the SARs of this series in more detail, we attempted to calculate logP (clogP) values for the acetamides 40–50 using the ACD/logP program.²¹ The clogP values obtained were compared with the CRAC channel inhibitory activity of the corresponding 5-(1-methyl-3-trifluoromethyl-1H-pyrazol-5-yl)-2-thiophenecarboxamides (4, 26–30, 32–35, 39, respectively), and the results are shown in Table 4. Among the acetamides bearing aromatic groups (40–47), the 4-chlorophenyl (40) and

 $^{^{}f}\%$ inhibition at 1 μ M.

g Not tested.

Table 4. $c \log P$ values of acetamides **40–50** and CRAC channel inhibitory activity of corresponding 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides

Compound	R	$c \log P^{a}$	Compound	CRAC IC ₅₀ ^b (μM)
40	4-Chlorophenyl	2.05	4	0.13
41	Thiophen-2-yl	1.43	26	0.10
49	Cyclohexyl	1.01	35	0.36
42	Thiophen-3-yl	0.76	27	0.61
43	1-Methylpyrrol-2-yl	0.74	28	0.58
45	Pyridin-4-yl	0.59	30	0.42
44	Pyridin-3-yl	0.26	29	0.99
48	Ethyl	-0.52	34	1.7
50	Piperidin-4-yl	-0.85	39	10% ^c
46	1,2,4-Triazol-4-yl	-1.43	32	6%°
47	Tetrazol-5-yl	-1.47	33	12% ^c

^a ACD/log P values.

thiophen-2-yl (41) derivatives gave $c \log P$ values greater than 1, and the corresponding compounds 4 and 26 were potent inhibitors of CRAC channels. In contrast, 1,2,4triazol-4-yl (46) and tetrazol-5-yl (47) groups produced hydrophilic acetamides and the corresponding thiophene-2-carboxamides 32 and 33 failed to inhibit CRAC channels. From these results, we hypothesized that a hydrophobic acetamide moiety would be an essential feature of potent inhibitors of CRAC channels. This speculation was supported by the fact that compounds 27–30, whose corresponding acetamides possessed $c \log P$ values of 0.2–0.8, showed moderate activity with IC₅₀ values of 0.4–1.0 μ M. In addition, the $c \log P$ value of the cyclohexyl derivative (49) was the highest in the N-aliphatic acetamide series 48–50, and the N-cyclohexylthiophene-2-carboxamide 35 was the most potent CRAC channel inhibitor of compounds 34, 35, and 39 (Table 4).

A selection of compounds which showed potent and selective inhibitory activity for CRAC channels (11, 12, 16, 17, 22-26, 28, 29, 35, and 36) was tested for their ability to inhibit phytohemagglutinin (PHA)-induced IL-2 production in Jurkat T lymphocytes⁷ (Table 5). Compound 4 inhibited IL-2 production with an IC₅₀ value of 0.53 μM. Substitutions at the 2-position of the phenyl group (22-25) increased the inhibitory activity, with compounds 23 and 25 in particular inhibiting IL-2 production with IC₅₀ values of 0.023 and 0.026 μ M, respectively. The 1-methylpyrrolyl derivative 28 was also a more potent inhibitor than compound 4 in this assay, despite having shown a 4-fold lower potency as a CRAC channel inhibitor. These results suggested that substitutions adjacent to the amide bond would be likely to increase the inhibitory effect of compounds on IL-2 production. The cyclohexyl derivative 35 inhibited IL-2 production with an IC₅₀ value of $0.076 \mu M$, which showed that hydrophobic groups in this moiety may be crucial for inhibitory activity against both IL-2 production and CRAC channels. Some compounds which

Table 5. Biological properties of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamides in vitro

Compound	IC ₅₀ (μM)		
	$\overline{\text{CRAC}^{\text{a}}}$	IL-2 ^b	
4	0.13	0.53	
11	0.14	0.75	
12	0.27	1.8	
16	0.062	0.85	
17	0.37	0.54	
36	0.24	1.5	
22	0.050	0.14	
23	0.09	0.023	
24	0.30	0.053	
25	0.14	0.026	
26	0.10	0.70	
28	0.58	0.20	
29	0.99	1.4	
35	0.36	0.076	

^a See the footnote a of Table 1.

were potent inhibitors of both CRAC channel and IL-2 production (16, 22–25, 35) were further tested for their ability to inhibit concanavalin A (Con A)-induced hepatitis in mice (Table 6).²² Among the compounds tested,

 Table 6. Effects of selected compounds on Con A-induced hepatitis in mice

Compound	In vivo hepatitis ^a		
	ED ₅₀ (mg/kg <i>p.o.</i>)	% inhibition at 30 mg/kg p.o.	
16		48	
22		46	
23		32	
24	12.7	67	
25		0	
35		9	

^a Inhibitory activities against Con A-induced liver injury in mice. See experimental section.

^b See the footnote a of Table 1.

c% inhibition at 10 μM.

^b Inhibition of PHA-induced IL-2 production in Jurkat T lymphocytes. See experimental section.

24 was found to be the most potent inhibitor with an ED₅₀ value of 12.7 mg/kg p.o.

4. Conclusion

We designed and synthesized novel 5-(1-methyl-3-trifluoromethyl-1H-pyrazol-5-yl)-2-thiophenecarboxamides based on compound 4 and evaluated their ability to inhibit CRAC and VOC channels. SAR study showed a relationship between the ability of compounds to inhibit CRAC channels and the $\sigma_{\rm para}$ values of the groups substituted at the 4-position on the phenyl ring, with values of around 0 being most favorable for a high potency. In addition, substitutions at the 2-position resulted in an improvement of the selectivity of the CRAC channel inhibitors and hydrophobic moieties such as chlorophenyl and cyclohexyl groups were shown to be essential to potent CRAC channel inhibitory activity. We identified compound 22 as the most promising novel CRAC channel inhibitor, with an IC₅₀ value of $0.050 \mu M$ and a selectivity index of more than 200. Additionally, some other compounds including compound 24 were found to inhibit PHA-induced IL-2 production in vitro with IC₅₀ values of the order of 10^{-8} M, and Con A-induced hepatitis in vivo, after oral administration. This study demonstrated that T lymphocyte function can be inhibited by CRAC channel inhibitors both in vitro and in vivo. These compounds have potential uses as a new class of orally active anti-inflammatory agents.

5. Experimental

5.1. Chemistry

Melting points were determined with a Yanaco MP-500D melting point apparatus and are uncorrected. ¹H NMR spectra were recorded on a JEOL JNM-EX90, JEOL JNM-LA300, JEOL JNM-EX400 or JEOL JNM-A500 spectrometer and were referenced to an internal standard, tetramethylsilane. The abbreviations used for the signal patterns are as follows: s, singlet; br s, broad singlet; d, doublet; t, triplet; q, quartet; dd, double doublet; dt, double triplet; tt, triple triplet; m, multiplet. Mass spectra were recorded on a Hitachi M-80 or JEOL JMS-DX300 mass spectrometer, and the ionization method was chosen from EI and FAB. High-resolution mass spectra (HRMS) were recorded on VG ZAB-VSE mass spectrometers. The elemental analyses were performed with a Yanako MT-5 microanalyzer (C, H, and N) and a Yokogawa IC-7000S ion chromatographic analyzer (halogens and S). Drying of organic solutions during workup was done over anhydrous MgSO₄. Preparative column chromatography was performed with Wakogel C-200 or Merck silica gel 60.

5.2. 1-Methyl-5-(2-thienyl)-3-trifluoromethyl-1*H*-pyrazole (7)

NaOMe (10.3 g, 191 mmol) was added to a solution of 2-acetylthiophene (5, 18.5 g, 147 mmol) in MeOH

(150 mL), and the whole mixture was stirred for 20 min at room temperature. It was cooled on ice-bath, then added ethyl trifluoroacetate (25.0 g, 176 mmol), and the whole mixture was heated to reflux for 19 h. H₂O (300 mL) was added to the mixture and extracted with AcOEt, washed with brine, dried, and concentrated in vacuo to give a pale brown solid (6, 34.7 g). This crude compound (6, 20.0 g) was added to a mixture of methylhydrazine (4.56 g, 99.0 mmol), AcOH (20 mL), and EtOH (200 mL), and the whole mixture was heated to reflux for 30 min. The mixture was concentrated in vacuo, and saturated aqueous NaHCO3 was added to the residue and extracted with AcOEt, washed with brine, dried, and concentrated in vacuo. The residue was purified by column chromatography (hexane/AcOEt, 20:1–10:1) to give 7 (11.8 g, 60%): ¹H NMR $(CDCl_3)$ δ 4.01 (3H, s), 6.64 (1H, s), 7.15 (1H, dd, J = 5.2, 3.7 Hz), 7.21 (1H, dd, J = 3.7, 1.3 Hz), 7.47 (1H, dd, J = 5.2, 1.3 Hz); FAB-MS $m/z 233 [(M+H)^{+}]$.

5.3. Ethyl 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxylate (8)

A 1.6 M solution of *n*-BuLi (33.0 mL, 52.8 mmol) in hexane was added dropwise to a solution of 1-methyl-5-(2-thienyl)-3-trifluoromethyl-1*H*-pyrazole (7, 11.2 g, 48.2 mmol) in THF (150 mL) at -70 °C. The mixture was stirred for 1.5 h at -70 °C and then ethyl chloroformate (10.5 g, 96.8 mmol) was added at the same temperature. The mixture was stirred for 15 min and then aqueous NH₄Cl was added. The whole mixture was extracted with AcOEt, washed with brine, dried, and concentrated in vacuo. The residue was purified by column chromatography (hexane/AcOEt, 7:1) to give **8** (9.58 g, 65%) as a pale yellow solid: ¹H NMR (CDCl₃) δ 1.40 (3H, t, J = 7.3 Hz), 4.05 (3H, s), 4.39 (2H, q, J = 7.0 Hz), 6.71 (1H, s), 7.20 (1H, d, J = 4.5 Hz), 7.80 (1H, d, J = 3.5 Hz); FAB-MS m/z 305 [(M+H)⁺].

5.4. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxylic acid (9)

A mixture of ethyl 5-(1-methyl-3-trifluoromethyl-1H-pyrazol-5-yl)-2-thiophenecarboxylate (**8**, 3.07 g, 10.1 mmol), 1 M NaOH (15 mL), and EtOH (30 mL) was stirred for 4 h at room temperature. One molar HCl (18 mL) was added to the mixture and extracted with AcOEt, washed with brine and dried, and then concentrated in vacuo to give **9** (2.79 g, 100%) as a pale brown solid: 1 H NMR (DMSO- d_{6}) δ 4.06 (3H, s), 7.18 (1H, s), 7.59 (1H, d, J = 3.9 Hz), 7.80 (1H, d, J = 3.9 Hz), 13.44 (1H, br s); FAB-MS m/z 275 [(M-H) $^{-}$].

5.5. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarbonylchloride (10)

A mixture of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxylic acid (9, 2.21 g, 8.00 mmol), oxalyl chloride (2.03 g, 16.0 mmol), and DMF (3 drops) in ClCH₂CH₂Cl (30 mL) was stirred for 90 min at room temperature. The mixture was concentrated in vacuo to give **10** (2.23 g, 95%) as a brown solid: ¹H NMR

(CDCl₃) δ 4.09 (3H, s), 6.78 (1H, s), 7.30 (1H, d, J = 4.4 Hz), 8.00 (1H, d, J = 3.9 Hz); FAB-MS m/z 291 $[(M+H)^{+}]$.

5.6. *N*-(4-Fluorophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (11)

To a mixture of 4-fluoroaniline (93 mg, 0.84 mmol), saturated aqueous NaHCO₃ (3 mL), and CH₂Cl₂ (2 mL), 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarbonylchloride (10, 195 mg, 0.662 mmol) was added and the whole mixture was stirred for overnight at room temperature. H₂O was added to the mixture and extracted with AcOEt, washed with brine and dried. and then concentrated in vacuo. The residue was recrystallized from AcOEt to give 11 (64 mg, 26%) as a colorless powder: mp 189–192 °C; ¹H NMR (CDCl₃) δ 4.07 (3H, s), 6.72 (1H, s), 7.08 (2H, t, J = 8.6 Hz), 7.23 (1H, d, J = 3.9 Hz), 7.55–7.61 (2H, m), 7.62 (1H, d, J = 3.9 Hz), 7.69 (1H, br s); FAB-MS m/z 370 $[(M+H)^{+}]$. Anal. Calcd for $C_{16}H_{11}F_{4}N_{3}OS$: C, 52.03; H, 3.00; N, 11.38; F, 20.58; S, 8.68. Found: C, 51.98; H, 2.84; N, 11.43; F, 20.75; S, 8.72.

The following compounds were prepared following the same method. Among the corresponding amines, 2-amino-1-methylpyrrole was prepared by the methods reported previously.²³

5.7. *N*-(4-Bromophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (12)

Yield 11%; mp 191–192 °C (AcOEt); ¹H NMR (CDCl₃) δ 4.07 (3H, s), 6.73 (1H, s), 7.24 (1H, d, J = 3.9 Hz), 7.47–7.55 (4H, m), 7.62 (1H, d, J = 3.9 Hz), 7.67 (1H, br s); FAB-MS m/z 430, 432 [(M+H) $^{+}$]. Anal. Calcd for C₁₆H₁₁BrF₃N₃OS: C, 44.67; H, 2.58; N, 9.77; Br, 18.57; F, 13.25; S, 7.45. Found: C, 44.56; H, 2.75; N, 9.80; Br, 18.62; F, 13.30; S, 7.36.

5.8. *N*-(4-Cyanophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (14)

Yield 55%; mp 211–214 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 4.09 (3H, s), 7.20 (1H, s), 7.67 (1H, d, J = 3.9 Hz), 7.85 (2H, d, J = 8.8 Hz), 7.95 (2H, d, J = 9.3 Hz), 8.15 (1H, d, J = 3.9 Hz), 10.72 (1H, s); FAB-MS m/z 377 [(M+H)⁺]. Anal. Calcd for C₁₇H₁₁F₃N₄OS · 0.15H₂O: C, 53.87; H, 3.00; N, 14.78; F, 15.04; S, 8.46. Found: C, 53.49; H, 2.85; N, 14.53; F, 15.41; S, 8.46.

5.9. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(4-nitrophenyl)-2-thiophenecarboxamide (15)

Yield 24%; mp 183–184 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 4.10 (3H, s), 7.20 (1H, s), 7.68 (1H, d, J = 3.9 Hz), 8.03 (2H, d, J = 9.3 Hz), 8.18 (1H, d, J = 4.4 Hz), 8.29 (2H, d, J = 9.2 Hz), 10.88 (1H, s); FAB-MS m/z 397 [(M+H)⁺]. HRMS m/z Calcd for $C_{16}H_{11}F_3N_4O_3S$ [(M+H)⁺]: 397.0582. Found: 397.0601.

5.10. *N*-(4-Isopropylphenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (18)

Yield 71%; mp 163–164 °C (AcOEt–hexane); 1 H NMR (DMSO- d_{6}) δ 1.21 (6H, d, J = 6.8 Hz), 2.83–2.92 (1H, m), 4.08 (3H, s), 7.17 (1H, s), 7.24 (2H, d, J = 8.3 Hz), 7.63 (1H, d, J = 3.9 Hz), 7.63 (2H, d, J = 8.8 Hz), 8.09 (1H, d, J = 3.9 Hz), 10.30 (1H, s); FAB-MS m/z 394 [(M+H) $^{+}$]. Anal. Calcd for C₁₉H₁₈F₃N₃OS: C, 58.00, H, 4.61; N, 10.68; F, 14.49; S, 8.15. Found: C, 58.17; H, 4.59; N, 10.68; F, 14.48; S, 8.16.

5.11. *N*-(4-Dimethylaminophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (20)

Yield 60%; mp 202–203 °C (EtOH–AcOEt); ¹H NMR (DMSO- d_6) δ 2.88 (6H, s), 4.08 (3H, s), 6.74 (2H, d, J = 9.2 Hz), 7.15 (1H, s), 7.52 (2H, d, J = 9.3 Hz), 7.60 (2H, d, J = 4.4 Hz), 8.04 (1H, d, J = 4.4 Hz), 10.13 (1H, s); FAB-MS m/z 394 (M⁺). Anal. Calcd for C₁₈H₁₇F₃N₄OS: C, 54.81, H, 4.34; N, 14.20; F, 14.34; S, 8.13. Found: C, 54.73; H, 4.43; N, 14.19; F, 14.34; S, 7.95.

5.12. *N*-(3-Chlorophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (21)

Yield 39%; mp 127–129 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 4.09 (3H, s), 7.17–7.21 (2H, m), 7.41 (1H, t, J = 8.1 Hz), 7.65 (1H, d, J = 3.9 Hz), 7.66–7.70 (1H, m), 7.91 (1H, t, J = 2.0 Hz), 8.11 (1H, d, J = 3.9 Hz), 10.51 (1H, s); FAB-MS m/z 386 [(M+H)⁺]. Anal. Calcd for C₁₆H₁₁ClF₃N₃OS: C, 49.81; H, 2.87; N, 10.89; Cl, 9.19; F, 14.77; S, 8.31. Found: C, 49.74; H, 2.73; N, 10.94; Cl, 9.11; F, 14.73; S, 8.29.

5.13. *N*-(1-Methyl-1*H*-pyrrol-2-yl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (28)

Yield 36%; mp 158–159 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 3.44 (3H, s), 4.07 (3H, s), 5.92 (1H, br s), 5.98 (1H, t, J = 3.2 Hz), 6.69 (1H, s), 7.16 (1H, s), 7.62 (1H, d, J = 3.9 Hz), 8.03 (1H, d, J = 3.9 Hz), 10.12 (1H, s); FAB-MS m/z 355 [(M+H) $^+$]. Anal. Calcd for C₁₅H₁₃F₃N₄OS: C, 50.84; H, 3.70; N, 15.81; F, 16.08; S, 9.05. Found: C, 50.90; H, 3.66; N, 15.77; F, 16.07; S, 9.01.

5.14. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(pyridin-4-yl)-2-thiophenecarboxamide monohydrochloride (30)

Yield 24%; mp 247 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 4.11 (3H, s), 7.25 (1H, s), 7.73 (1H, d, J=4.3 Hz), 8.39 (2H, d, J=5.9 Hz), 8.53 (1H, d, J=3.9 Hz), 8.76 (2H, d, J=6.4 Hz), 11.93 (1H, s); FAB-MS m/z 353 [(M+H)⁺]. Anal. Calcd for C₁₅H₁₁F₃N₄OS · HCl: C, 46.34; H, 3.11; N, 14.41; Cl, 9.12; F, 14.66; S, 8.25. Found: C, 46.19; H, 2.99; N, 14.39; Cl, 9.07; F, 14.72; S, 8.13.

5.15. *N*-Cyclohexyl-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (35)

Yield 68%; mp 140–141 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 1.10–1.20 (1H, m), 1.24–1.36 (4H, m), 1.58–1.66 (1H, m), 1.69–1.87 (4H, m), 3.67–3.77 (1H, m), 4.05 (3H, s), 7.10 (1H, s), 7.53 (1H, d, J = 3.9 Hz), 7.87 (1H, d, J = 3.9 Hz), 8.38 (1H, d, J = 7.8 Hz); FAB-MS m/z 358 [(M+H) $^+$]. Anal. Calcd for C₁₆H₁₈F₃N₃OS: C, 53.77; H, 5.08; N, 11.76; F, 15.95; S, 8.97. Found: C, 53.47; H, 4.94; N, 11.68; F, 16.25; S, 8.97.

The following compounds were prepared according to this method, substituting Et₃N for aqueous NaHCO₃. Among the corresponding amines, 2- and 3-aminothiophene were prepared by the methods reported previously.²⁴

5.16. *N*-(4-Methylphenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (17)

Yield 61%; mp 167–169 °C (AcOEt–hexane); ¹H NMR (CDCl₃) δ 2.35 (3H, s), 4.06 (3H, s), 6.72 (1H, s), 7.19 (2H, d, J = 8.3 Hz), 7.22 (1H, d, J = 3.9 Hz), 7.49 (2H, d, J = 8.3 Hz), 7.60 (1H, d, J = 3.9 Hz), 7.65 (1H, br s); FAB-MS m/z 366 [(M+H)⁺]. Anal. Calcd for C₁₇H₁₄F₃N₃OS · 0.25H₂O: C, 55.20; H, 3.95; N, 11.36; F, 15.41; S, 8.67. Found: C, 55.15; H, 3.77; N, 11.43; F, 15.50; S, 8.87.

5.17. *N*-(2-Fluorophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (23)

Yield 26%; mp 147–148 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 4.09 (3H, s), 7.18 (1H, s), 7.22–7.35 (3H, m), 7.58–7.62 (1H, m), 7.64 (1H, d, J = 3.9 Hz), 8.10 (1H, d, J = 3.9 Hz), 10.33 (1H, s); FAB-MS m/z 370 [(M+H)⁺]. Anal. Calcd for C₁₆H₁₁F₄N₃OS: C, 52.03; H, 3.00; N, 11.38; F, 20.58; S, 8.68. Found: C, 51.81; H, 2.92; N, 11.41; F, 20.55; S, 8.69.

5.18. *N*-(2-Methylphenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (24)

Yield 50%; mp 159–161 °C (AcOEt–hexane); ¹H NMR (CDCl₃) δ 2.35 (3H, s), 4.06 (3H, s), 6.72 (1H, s), 7.13–7.18 (1H, m), 7.22 (1H, d, J = 3.9 Hz), 7.23–7.30 (2H, m), 7.58 (1H, s), 7.61 (1H, d, J = 3.9 Hz), 7.86 (1H, d, J = 7.8 Hz); FAB-MS m/z 366 [(M+H)⁺]. Anal. Calcd for C₁₇H₁₄F₃N₃OS: C, 55.88; H, 3.86; N, 11.50; F, 15.60; S, 8.78. Found: C, 55.77; H, 3.79; N, 11.64; F, 15.56; S, 8.77.

5.19. *N*-(2,4-Dichlorophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (25)

Yield 25%; mp 181–183 °C (AcOEt–hexane); ¹H NMR (CDCl₃) δ 4.08 (3H, s), 6.74 (1H, s), 7.26 (1H, d, J = 3.9 Hz), 7.32 (1H, dd, J = 9.1, 2.2 Hz), 7.45 (1H, d, J = 2.4 Hz), 7.66 (1H, d, J = 3.9 Hz), 8.25 (1H, s), 8.45 (1H, d, J = 8.8 Hz); FAB-MS m/z 420 [(M+H)⁺]. Anal. Calcd for C₁₆H₁₀Cl₂F₃N₃OS: C, 45.73; H, 2.40; N,

10.00; Cl, 16.87; F, 13.56; S, 7.63. Found: C, 45.65; H, 2.38; N, 9.99; Cl, 16.77; F, 13.40; S, 7.59.

5.20. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(thiophen-2-yl)-2-thiophenecarboxamide (26)

Yield 15%; mp 180–185 °C (Et₂O–hexane); ¹H NMR (CDCl₃) δ 4.07 (3H, s), 6.73 (1H, s), 6.83 (1H, dd, J = 3.7, 1.2 Hz), 6.92 (1H, dd, J = 5.7, 3.7 Hz), 6.97–7.00 (1H, m), 7.24 (1H, d, J = 3.9 Hz), 7.65 (1H, d, J = 3.9 Hz), 8.37 (1H, br s); FAB-MS m/z 358 [(M+H)⁺]. Anal. Calcd for C₁₄H₁₀F₃N₃OS₂ · 0.25H₂O: C, 46.47; H, 2.92; N, 11.61; F, 15.75; S, 17.72. Found: C, 46.37; H, 2.71; N, 11.44; F, 15.65; S, 17.72.

5.21. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(thiophen-3-yl)-2-thiophenecarboxamide (27)

Yield 62%; mp 166–168 °C (AcOEt–hexane); ¹H NMR (CDCl₃) δ 4.07 (3H, s), 6.72 (1H, s), 7.13 (1H, dd, J = 4.9, 1.5 Hz), 7.23 (1H, d, J = 3.9 Hz), 7.31 (1H, dd, J = 4.7, 3.2 Hz), 7.60 (1H, d, J = 3.9 Hz), 7.68 (1H, dd, J = 3.0, 1.4 Hz), 7.97 (1H, s); FAB-MS mlz 358 [(M+H)⁺]. Anal. Calcd for C₁₄H₁₀F₃N₃OS₂: C, 47.05; H, 2.82; N, 11.76; F, 15.95; S, 17.95. Found: C, 46.91; H, 2.83; N, 11.68; F, 16.24; S, 17.91.

5.22. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(pyridin-3-yl)-2-thiophenecarboxamide (29)

Yield 62%; mp 186–187 °C (AcOEt–hexane); ¹H NMR (CDCl₃) δ 4.07 (3H, s), 6.73 (1H, s), 7.25 (1H, d, J = 3.9 Hz), 7.35 (1H, dd, J = 8.3, 3.8 Hz), 7.68 (1H, d, J = 3.9 Hz), 7.95 (1H, s), 8.24–8.28 (1H, m), 8.42 (1H, dd, J = 4.7, 1.2 Hz), 8.68 (1H, d, J = 2.5 Hz); FAB-MS m/z 353 [(M+H)⁺]. Anal. Calcd for C₁₅H₁₁F₃N₄OS: C, 51.13; H, 3.15; N, 15.90; F, 16.18; S, 9.10. Found: C, 51.03; H, 3.14; N, 15.84; F, 15.89; S, 9.04.

5.23. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(1*H*-tetrazol-5-yl)-2-thiophenecarboxamide (33)

Yield 72%; mp 285–287 °C (acetone); ¹H NMR (DMSO- d_6) δ 4.09 (3H, s), 7.22 (1H, s), 7.69 (1H, d, J = 3.9 Hz), 8.32 (1H, d, J = 4.4 Hz), 12.67 (1H, br s); FAB-MS m/z 344 [(M+H)⁺]. Anal. Calcd for C₁₁H₈F₃N₇OS · 0.2H₂O: C, 38.09; H, 2.44; N, 28.26; F, 16.43; S, 9.24. Found: C, 37.95; H, 2.40; N, 28.54; F, 16.78; S, 9.24.

The following compounds were prepared according to this method, substituting pyridine for aqueous NaHCO₃.

5.24. Ethyl 4-{[5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarbonyllamino}benzoate (13)

Yield 45%; mp 183–185 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 1.33 (3H, t, J = 7.1 Hz), 4.09 (3H, s), 4.31 (2H, q, J = 7.2 Hz), 7.19 (1H, s), 7.66 (1H, d, J = 3.9 Hz), 7.91 (2H, d, J = 8.7 Hz), 7.99 (2H, d, J = 8.8 Hz), 8.16 (1H, d, J = 4.4 Hz), 10.65 (1H, s); FAB-MS m/z 424 [(M+H)⁺]. Anal. Calcd for

C₁₉H₁₆F₃N₃O₃S: C, 53.90; H, 3.81; N, 9.92; F, 13.46; S, 7.57. Found: C, 53.89; H, 3.79; N, 9.87; F, 13.43; S, 7.48.

5.25. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-phenylthiophene-2-carboxamide (16)

Yield 86%; mp 129–130 °C; ¹H NMR (CDCl₃) δ 4.07 (3H, s), 6.72 (1H, s), 7.19 (1H, t, J = 7.6 Hz), 7.23 (1H, d, J = 3.4 Hz), 7.40 (2H, t, J = 8.1 Hz), 7.60–7.63 (3H, m), 7.66 (1H, br s); FAB-MS m/z 352 [(M+H)⁺]. Anal. Calcd for C₁₆H₁₂F₃N₃OS: C, 54.70; H, 3.44; N, 11.96; F, 16.22; S, 9.13. Found: C, 54.69; H, 3.22; N, 12.21; F, 16.10; S, 9.05.

5.26. *N*-(4-Methoxyphenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (19)

Yield 49%; mp 166–167 °C (AcOEt–hexane); ¹H NMR (DMSO- d_6) δ 3.75 (3H, s), 4.08 (3H, s), 6.95 (2H, d, J = 9.3 Hz), 7.16 (1H, s), 7.62 (1H, d, J = 3.9 Hz), 7.62 (2H, d, J = 9.3 Hz), 8.06 (1H, d, J = 3.9 Hz), 10.27 (1H, s); FAB-MS m/z 382 [(M+H)⁺]. Anal. Calcd for C₁₇H₁₄F₃N₃O₂S: C, 53.54, H, 3.70; N, 11.02; F, 14.94; S, 8.41. Found: C, 53.45; H, 3.67; N, 10.96; F, 14.88; S, 8.28.

5.27. N-(2-Chlorophenyl)-5-(1-methyl-3-trifluoromethyl-1H-pyrazol-5-yl)-2-thiophenecarboxamide (22)

Yield 41%; mp 156 °C (EtOH); 1 H NMR (CDCl₃) δ 4.08 (3H, s), 6.74 (1H, s), 7.11 (1H, td, J = 7.8, 1.5 Hz), 7.26 (1H, d, J = 3.4 Hz), 7.35 (1H, td, J = 7.8, 1.4 Hz), 7.44 (1H, dd, J = 7.8, 1.5 Hz), 7.66 (1H, d, J = 3.9 Hz), 8.32 (1H, s), 8.48 (1H, dd, J = 8.3, 1.4 Hz); FAB-MS m/z 386 [(M+H) $^{+}$]. Anal. Calcd for C₁₆H₁₁ClF₃N₃OS: C, 49.81; H, 2.87; N, 10.89; Cl, 9.19; F, 14.77; S, 8.31. Found: C, 49.79; H, 2.80; N, 10.95; Cl, 9.18; F, 14.73; S, 8.30.

5.28. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(thiazol-2-yl)-2-thiophenecarboxamide (31)

Yield 58%; mp 235–238 °C (EtOH); ¹H NMR (DMSO- d_6) δ 4.09 (3H,s), 7.20 (1H, s), 7.29 (1H, d, J = 3.4 Hz), 7.57 (1H, d, J = 3.9 Hz), 7.65 (1H, d, J = 3.9 Hz), 8.28 (1H, br s), 12.94 (1H, br s); FAB-MS m/z 359 [(M+H)⁺]. Anal. Calcd for C₁₃H₉F₃N₄OS₂: C, 43.57; H, 2.53; N, 15.63; F, 15.90; S, 17.90. Found: C, 43.34; H, 2.48; N, 15.64; F, 16.20; S, 17.82.

5.29. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(4*H*-1,2,4-triazol-4-yl)-2-thiophenecarboxamide (32)

Yield 71%; mp 207–210 °C (AcOEt); ¹H NMR (DMSO- d_6) δ 4.08 (3H, s), 7.22 (1H, s), 7.69 (1H, d, J = 3.9 Hz), 7.98 (1H, d, J = 3.9 Hz), 8.83 (2H, s), 12.34 (1H, s); FAB-MS m/z 343 [(M+H)⁺]. Anal. Calcd for C₁₂H₉F₃N₆OS: C, 42.11; H, 2.65; N, 24.55; F, 16.65; S, 9.37. Found: C, 41.77; H, 2.55; N, 24.20; F, 16.59; S, 9.17.

5.30. *N*-Ethyl-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (34)

To a mixture of ethylamine (70% in H₂O, 1 mL, 15.5 mmol) and THF (4 mL), 5-(1-methyl-3-trifluoro-

methyl-1*H*-pyrazol-5-yl)-2-thiophenecarbonylchloride (**10**, 150 mg, 0.509 mmol) was added and the whole mixture was stirred for 2 h at room temperature. H₂O was added to the mixture and extracted with AcOEt, washed with brine and dried, and then concentrated in vacuo. The residue was recrystallized from AcOEt–hexane to give **34** (96 mg, 62%) as a colorless powder: mp 136–137 °C; ¹H NMR (DMSO- d_6) δ 1.14 (3H, t, J = 7.1 Hz), 3.25–3.32 (2H, m), 4.05 (3H, s), 7.11 (1H, s), 7.54 (d, J = 3.9 Hz), 7.81 (1H, d, J = 3.9 Hz), 8.65 (1H, t, J = 5.4 Hz); FAB-MS m/z 304 [(M+H)⁺]. Anal. Calcd for C₁₂H₁₂F₃N₃OS: C, 47.52; H, 3.99; N, 13.85; F, 18.79; S, 10.57. Found: C, 47.45; H, 3.98; N, 13.83; F, 19.07; S, 10.46.

5.31. *N*-(4-Hydroxyphenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (36)

A mixture of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophene (with Standard) carboxylic acid (9, 156 mg, 0.565 mmol), 4-aminophenol (62 mg, 0.57 mmol), and EDC · HCl (114 mg, 0.595 mmol) in THF (2 mL) was stirred for 3 days at room temperature. H₂O (10 mL) was added to the mixture and extracted with AcOEt, washed with 1 M HCl, saturated aqueous NaH-CO₃, brine and dried, and then concentrated in vacuo. The residue was purified by column chromatography (hexane/AcOEt, 2:1) and recrystallized from AcOEthexane to give 36 (113 mg, 54%) as a colorless powder: mp 185–186 °C; ¹H NMR (DMSO- d_6) δ 4.08 (3H, s), 6.76 (2H, d, J = 8.8 Hz), 7.15 (1H, s), 7.48 (2H, d, J = 9.3 Hz), 7.61 (1H, d, J = 4.4 Hz), 8.04 (1H, d, J = 3.9 Hz), 9.32 (1H, s), 10.17 (1H, s); FAB-MS m/z368 $[(M+H)^{+}]$. Anal. Calcd for $C_{16}H_{12}F_{3}N_{3}O_{2}S$: C, 52.31, H, 3.29; N, 11.44; F, 15.52; S, 8.73. Found: C, 51.99; H, 3.37; N, 11.27; F, 15.76; S, 8.68.

The following compound was prepared using similar method.

5.32. *tert*-Butyl 4-{[5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarbonyl]amino}piperidine-1-carboxylate (37)

¹H NMR (CDCl₃) δ 1.44–1.49 (2H, m), 1.47 (9H, s), 1.98–2.08 (2H, m), 2.83–2.97 (2H, m), 4.04 (3H, s), 4.08–4.17 (3H, m), 6.02–6.08 (1H, m), 6.68 (1H, s), 7.17 (1H, d, J = 3.6 Hz), 7.50 (1H, d, J = 3.9 Hz); FAB-MS m/z 459 [(M+H)⁺].

5.33. *N*-(4-Aminophenyl)-5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thiophenecarboxamide (38)

Zinc powder (240 mg, 3.67 mmol) and NH₄Cl (196 mg, 3.66 mmol) was added to a solution of 5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(4-nitrophenyl)-2-thiophenecarboxamide (**15**, 146 mg, 0.368 mmol) in EtOH–H₂O (2:1, 6 mL) at 5 °C. The mixture was stirred for 20 min at room temperature and filtered on Celite. H₂O was added to the filtrate and extracted with AcOEt, washed with 0.1 M HCl, saturated aqueous NaHCO₃, brine and dried, and then concentrated in vacuo. The residue was recrystallized from AcOEt–hexane to give

38 (64 mg, 47%) as a yellow powder: mp 181–183 °C; 1 H NMR (DMSO- d_{6}) δ 4.07 (3H, s), 4.99 (2H, s), 6.56 (2H, d, J = 8.8 Hz), 7.14 (1H, s), 7.32 (2H, d, J = 8.8 Hz), 7.59 (1H, d, J = 3.9 Hz), 8.01 (1H, d, J = 3.9 Hz), 10.02 (1H, s); FAB-MS m/z 367 [(M+H) $^{+}$]. Anal. Calcd for C₁₆H₁₃F₃N₄OS: C, 52.45, H, 3.58; N, 15.29; F, 15.56; S, 8.75. Found: C, 52.65; H, 3.47; N, 15.04; F, 15.35; S, 8.60.

5.34. 5-(1-Methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-*N*-(4-piperidyl)-2-thiophenecarboxamide monohydrochloride (39)

A mixture of tert-butyl 4-{[5-(1-methyl-3-trifluoromethyl-1*H*-pyrazol-5-yl)-2-thenoyl]amino}piperidine-1-carboxylate (37, 65 mg, 0.142 mmol) and 4 M HCl-AcOEt (1 mL) in AcOEt (1 mL) was stirred for 4 h at room temperature. The mixture was concentrated in vacuo, and the residue was recrystallized from EtOH to give 39 (34 mg, 61%) as a colorless powder: mp 265–266 °C; ¹H NMR (DMSO- d_6) δ 1.73–1.85 (2H, m), 1.93–2.01 (2H, m), 2.96–3.06 (2H, m), 3.28–3.32 (2H, m), 4.00– 4.08 (1H, m), 4.05 (3H, s), 7.12 (1H, s), 7.56 (1H, d, J = 3.9 Hz), 7.97 (1H, d, J = 3.9 Hz), 8.74 (1H, d, J = 7.4 Hz), 8.78–8.90 (1H, br s); FAB-MS m/z 359 $[(M+H)^{+}]$. Anal. Calcd for $C_{15}H_{17}F_{3}N_{4}OS \cdot HCl$: C, 45.63; H, 4.59; N, 14.19; Cl, 8.98; F, 14.43; S, 8.12. Found: C, 45.32; H, 4.76; N, 14.20; Cl, 8.89; F, 14.63; S, 8.10.

5.35. QSAR analysis

Linear and multiple regression analyses were carried out using TSAR 3.0 (Oxford Molecular Group). One or two out of the following parameters (π , molecular refractivity, Swein and Lupton F/R, σ_{para} (σ_{para}^2), E_s , Verloop L, molecular volume, molecular surface area, and Verloop B5) were applied to the analyses. The following statistical parameters were determined for each regression equation: the number of points: n; the correlation coefficient: r; the significance of the regression model: F; the standard error: s.

5.36. Fura-2 loading and population intracellular calcium measurements

Cells were suspended in Hepes buffered solution (pH 7.4) of the following composition: NaCl: 137 mM, KCl: 5.8 mM, MgCl₂: 1 mM, CaCl₂: 2.5 mM, glucose: 5 mM, and Hepes: 10 mM. The cells were loaded with 1 μM Fura-2/AM at room temperature for 45 min, followed by successive washes to remove unincorporated dye, and resuspended in Hepes buffered solution. Cell suspensions were studied in a 96-well black plate. Fluorescence measurements for the determination of intracalcium concentration were fluorescence microplate reader (Fluostar, SLT Labinstruments Ges.m.b.H, Austria) with excitation wavelength of each 340 and 380 nm at emission fluorescence detection of 500 nm. Then, self-fluorescence of each compound was measured in a cell-free way and was deducted from cell way fluorescence. Final intracellular calcium concentrations in each well were calculated from the fluorescence ratio, using the standard equation. The $R_{\rm max}$ value was obtained from 25 μ M ionomycintreated wells. The $R_{\rm min}$ value was obtained from 25 μ M ionomycin/50 mM EGTA-treated wells.

5.37. CRAC channel inhibition assay

CRAC channel inhibition was evaluated in Jurkat cells $(2 \times 10^6 \text{ cells/mL})$. Fura-2 loaded Jurkat cells were stimulated with 1 μ M thapsigargin for 30 min, and measured intracellular calcium concentration at the endpoint of 30 min. IC₅₀ values on CRAC channel inhibition of each compounds were calculated from percent inhibition of thapsigargin-induced calcium influx in Jurkat cells.

5.38. VOC channel inhibition assay

VOC channel inhibition was evaluated in murine neuroblastoma, PC12-h5 cells (1×10^6 cells/mL). Fura-2 loaded PC12-h5 cells were stimulated with 50 mM KCl for 20 min, and measured intracellular calcium concentration at the endpoint of 20 min. IC₅₀ values on VOC channel inhibition of each compounds were calculated from percent inhibition of KCl-induced calcium influx in PC12 cells.

5.39. IL-2 production assay

Jurkat T lymphocytes $(5 \times 10^6 \text{ cells/mL})$ were placed in a 96-well microplate and incubated with PHA (20 µg/mL) for 20 h and supernatant was collected from these cells. IL-2 concentration in each supernatant was measured by human IL-2 ELISA system (DuoSeTTM, Genzyme).

5.40. Con A-induced hepatitis

Con A (20 mg/kg) was intravenously injected to female Balb/c mice in a volume of 10 mL/kg. Blood samples were obtained 24 h after Con A injection, and serum GOT and GPT levels were measured. For oral dosage groups, compounds were suspended with 0.5% MC and administered orally in a volume of 10 mL/kg 1 h before Con A injection. Cyclosporin A (CsA) was dissolved in physiological saline and subcutaneously administered at 50 mg/kg in a volume of 10 mL/kg 2 h before Con A injection.

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References and notes

(a) Mori, Y.; Mikala, G.; Varadi, G.; Kobayashi, T.; Koch, S.; Wakamori, M.; Schwartz, A. *Jpn. J. Pharmacol.* 1996, 72, 83; For example: (b) Takami, T.; Shigemasa, M. *Hypertens. Res.* 2003, 26, 609; For example: (c) Jouvenceau, A.; Eunson, L. H.; Spauschus, A.; Ramesh, V.; Zuberi, S. M.; Kullmann, D. M.; Hanna, M. G. *Lancet* 2001, 358, 801.

- (a) Hoth, M. Pflügers Arch. 1995, 430, 315; (b) Hoth, M.; Penner, R. Nature 1992, 355, 353; (c) Lewis, R. S.; Cahalan, M. D. Annu. Rev. Immunol. 1995, 13, 623; (d) Clementi, E.; Meldolesi, J. Cell Calcium 1996, 19, 269; (e) Geiszt, M.; Kapus, A.; Nemet, K.; Farkas, L.; Ligeti, E. J. Biol. Chem. 1997, 272, 26471.
- (a) Clapham, D. E. *Cell* 1995, 80, 259; (b) Petersen, O. H.; Gerasimenko, O. V.; Gerasimenko, J. V.; Mogami, H.; Tepikin, A. V. *Cell Calcium* 1998, 23, 87.
- 4. Berridge, M. J.; Irvine, R. F. Nature 1989, 341, 197.
- (a) Putney, J. W., Jr. Cell Calcium 1990, 11, 611; (b) Randriamampita, C.; Tsien, R. Y. Nature 1993, 364, 809; (c) Fasolato, C.; Innocenti, B.; Pozzan, T. Trends Pharmacol. Sci. 1994, 15, 77; (d) Berridge, J. M. Biochem. J. 1995, 312, 1; (e) Fanger, C. M.; Hoth, M.; Crabtree, G. R.; Lewis, R. S. J. Cell Biol. 1995, 131, 655; (f) Zweifach, A.; Lewis, R. S. J. Biol. Chem. 1995, 270, 14445; (g) Zweifach, A.; Lewis, R. S. J. Gen. Physiol. 1996, 107, 597.
- (a) Slaughter, R. S.; Garcia, M. L.; Kaczorowski, G. J. *Curr. Pharm. Design* **1996**, 2, 610; (b) Cahalan, M. D.; Chandy, K. G. *Curr. Opin. Biotechnol.* **1997**, 8, 749.
- Chung, S. C.; McDonald, T. V.; Gardner, P. Br. J. Pharmacol. 1994, 113, 861.
- Encabo, A.; Romanin, C.; Birke, F. W.; Kukovets, W. R.; Groschner, K. Br. J. Pharmacol. 1996, 119, 702.
- Felder, C. C.; Ma, A. L.; Liotta, L. A.; Kohn, E. C. J. Pharm. Exp. Ther. 1991, 257, 967.
- Kubota H., Yonetoku Y., Sugasawa K., Funatsu M., Kawazoe S., Toyoshima A., Okamoto Y., Ishikawa J., Takeuchi M. WO 9919303; Chem. Abstr. 1999, 130, 311815.
- 11. Reid, J. C.; Calvin, M. J. Am. Chem. Soc. 1950, 72, 2948.
- Mach, R. H.; Leudtke, R. R.; Unsworth, C. D.; Boundy, V. A.; Nowak, P. A.; Scripko, J. G.; Elder, S. T.; Jackson, J. R.; Hoffman, P. L.; Evora, P. H.; Rao, A. V.; Molinoff, P. B.; Childers, S. R.; Ehrenkaufer, R. L. J. Med. Chem. 1993, 36, 3707.

- (a) Ref. 7; (b) Timmerman, L. A.; Clipstone, N. A.; Ho, S. N.; Northrop, J. P.; Crabtree, G. R. Nature 1996, 383, 837.
- Virgilio, F. D.; Milani, D.; Leon, A.; Meldolesi, J.; Pozzan, T. J. Biol. Chem. 1987, 262, 9189.
- (a) Fujita, T.; Iwasa, J.; Hansch, C. J. Am. Chem. Soc. 1964, 86, 5175; (b) Iwasa, J.; Fujita, T.; Hansch, C. J. Med. Chem. 1965, 8, 153.
- 16. Hansch, C.; Calef, D. F. J. Org. Chem. 1976, 41, 1240.
- (a) Swain, C. G.; Lupton, E. C. J. Am. Chem. Soc. 1968,
 90, 4328; (b) Hansch, C.; Leo, A.; Uner, S. H.; Kim, K. H.; Nikaidani, D.; Lien, E. J. J. Med. Chem. 1973, 16, 1207.
- (a) Chapman N. B., Shorter J., Eds.; Advances in Linear Free Energy Relationship, Plenium Press: London, 1972.;
 (b) Chapman N. B., Shorter J., Eds.; Correlation Analysis in Chemistry, Plenium Press: London, 1978.
- (a) Taft, R. W. In Steric Effects in Organic Chemistry;
 Newman, M. S., Ed.; Wiley: New York, 1956; p 556; (b)
 Kutter, E.; Hansch, C. J. Med. Chem. 1969, 12, 647; (c)
 Fujita, T. J. Med. Chem. 1973, 16, 923.
- (a) Verloop A., Hoogenstraaten W., Tipker J. In Drug Design, Ariens, E. A., Ed.; Academic Press: New York, 1976; Vol. 7, pp 165.(b) Verloop A. Pesticide Chemistry, In Human Welfare and the Environment, Miyamoto, J., Kearney, P. C., Eds.; Pergamon Press: Oxford, 1983; Vol. 1, pp 339.; (c) Iwamura H. In Kozokassei Sokan to Drug Design, Fujita, T., Ed.; Kagaku Zokan, Kagaku Dojin: Kyoto, 1986; Vol. 107, pp 79.
- 21. ACD/log *P* DB program, ver. 3.5 for Microsoft Windows, LA systems Corp.
- Tiegs, G.; Hentschel, J.; Wendel, A. J. Clin. Invest. 1992, 90, 196.
- 23. De Rosa, M.; Issac, R. P.; Houghton, G. *Tetrahedron Lett.* **1995**, *36*, 9261.
- 24. Binder, D.; Habison, G.; Noe, C. R. Synthesis 1977, 4, 255